

## Review Article

# Proteinogenic Amino Acids Labelled with $^{15}\text{N}$ and/or $^{13}\text{C}$ for Application in Peptide Synthesis: A Short Review with a Comprehensive List of Published Derivatives

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**Abstract:** A review is given of the literature dealing with the most common protected derivatives of  $^{15}\text{N}$ - and/or  $^{13}\text{C}$ -labelled amino acids of interest in peptide synthesis. The list contains all such Boc-, Z- and Fmoc-amino acids as well as published methyl, ethyl, *t*-butyl and benzyl esters.

**Keywords:** Carbon-13; isotope-labelled amino acid; nitrogen-15; proteinogenic amino acid; stable isotope

## INTRODUCTION

Amino acids labelled with the stable nuclei  $^{15}\text{N}$  and/or  $^{13}\text{C}$  have played important roles in various metabolic, biosynthetic and mechanistic studies during recent decades. Since the pioneering synthetic experiments were performed in this field [1-4], altogether over 150 isotopomers of the 20 species commonly found in proteins have been described. More recently, such amino acids have also been incorporated into proteins for use in structural studies by various NMR techniques [5]. Although the natural abundance of the  $^{13}\text{C}$  nucleus is high enough (1.1%) to be useful in many cases without labelling, relatively large amounts of sample and long spectral accumulation times are required. The even lower natural abundance of  $^{15}\text{N}$  (0.37%) further stresses the need for labelling in order to exploit this nucleus in such studies.

For the synthesis of isotopically labelled peptides for structural and other purposes, the corresponding

protected amino acid derivatives are required. To date a considerable number of such have been described in the literature, most of which are not easy to retrieve without a systematic search. Therefore, these compounds are summarized for the first time in this review.

## ORGANIZATION OF THE REVIEW

This review is restricted entirely to the proteinogenic amino acids. It comprises only the most frequently applied protected derivatives used in peptide synthesis. These include the Boc-, Z- and Fmoc-amino protected derivatives, the methyl, ethyl, *t*-butyl and benzyl esters and the *t*-butyl and benzyl ethers and thioethers. For these compounds the *Formula Index of Chemical Abstracts* has been systematically searched up to June 1994. A few other protected derivatives found during this search have been added. All of these compounds are collected in Table 1.

Table 1 is arranged alphabetically with respect to the involved amino acid derivatives. Both uniformly

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and specifically labelled isotopomers are listed and to identify them unambiguously their Chemical Abstracts Service (CAS)-registry numbers are also included, together with those of the labelled parent free amino acids. In most cases, some information about the preparation or origin of the compounds and other relevant comments is also given. Finally, a reference is given to the original literature.

The methods of preparation used reflect the development of both amino acid synthesis and protection during the last decades. Free amino acids prepared chemically as racemates and resolved by use of enzymes are coded as A. Others of unspecified or commercial origin are characterized by B or C. The use of microbiological or biosynthetic methods is denoted by D. Conversion of an amino acid or derivative thereof into another labelled one without resolution is coded E. Asymmetric or other direct synthesis is indicated by F or G, respectively. Non-routine derivatization is generally pointed out under comments.

## COMMENTS

As shown in Table 1, nearly 100 isotopomers of derivatives of proteinogenic amino acids with relevance in peptide synthesis have been described in the literature to date. From them a considerable number of labelled peptides have been prepared and studied.

Further inspection of Table 1 demonstrates that no labelled derivatives with application in peptide synthesis have so far been prepared from the eight amino acids cystine, glutamic acid, histidine, isoleucine, lysine, methionine, threonine and tryptophan. For the remaining 11, excluding glycine, for which a large number of relevant species are available, an average of 5 different labelled protected derivatives that are of interest in this context have been made. Selectively  $^{15}\text{N}$ -labelled derivatives exist for alanine, glycine, leucine, phenylalanine, tyrosine and valine only. Such can nowadays generally be made from commercial D-amino acids via the corresponding hydroxy acids [6,39]. For a trifunc-

Table 1 List of Amino Acid Derivatives, Labelled with  $^{15}\text{N}$  and/or  $^{13}\text{C}$

No.	Derivative/pattern of labelling	CAS registry no./do. free amino acid	Preparation method	Comments	Ref.(s)
1	Boc-Ala $^{15}\text{N}$	139952-87-7 25713-23-9	E	Simultaneous protection	6
2	Boc-Ala $^{15}\text{N}, 1, 2-^{13}\text{C}_2$	---	F		7
3	Boc-Ala $^{15}\text{N}, 1, 2-^{13}\text{C}_2,$ $3, 3, 3-^{\text{D}}_3$	---	F		7
4	Boc-Ala U- $^{13}\text{C}$	72634-72-1 100108-77-8	C	90% enriched	8
5	Z-Ala $3-^{13}\text{C}$	155256-73-8 65163-25-9	B		9
6	Fmoc-Ala $^{15}\text{N}$	117398-49-9 25713-23-9	C		10
7	Ala-OMe·HCl $^{15}\text{N}$	75531-21-4 25713-23-9	B	ca. 1% $^{15}\text{N}$	11
8	Ala-OMe·HCl $1-^{13}\text{C}$	137295-73-9 21764-56-7	B		12
9	Ala-OBzl·Tos $^{15}\text{N}$	139929-02-5 25713-23-9	E		6
10	Z-Arg $1-^{13}\text{C}$	141074-93-3 81201-96-9	C		13
11	Z-Arg(Pmc) $1-^{13}\text{C}$	141074-94-4 81201-96-9	C		13
12	Fmoc-Arg(Pmc) $1-^{13}\text{C}$	141074-96-6 81201-96-9	C		13
13	Arg(Pmc) $1-^{13}\text{C}$	141074-95-5 81201-96-9	C		13

(continued)

Table 1 continued

No.	Derivative/pattern of labelling	CAS registry no./do. free amino acid	Preparation method	Comments	Ref.(s)
14	Boc-Asn 4- <sup>15</sup> N	150464-76-9 89695-60-3	E		14
15	Boc-Asn-OBzl 4- <sup>15</sup> N	150464-74-7 89695-60-3	E		14
16	Boc-Asp(OBzl) U- <sup>13</sup> C	72015-63-5 55443-54-4	D	85% enriched	15
17	Boc-Asp-(OBzl)-OSU U- <sup>13</sup> C	72015-64-6 55443-54-4	D	85% enriched	15
18	Z-Asp U- <sup>13</sup> C	86418-73-7 55443-54-4	D	85% enriched	16
19	Fmoc-Asp(O <sup>t</sup> Bu) 3- <sup>13</sup> C	141074-92-2 68261-19-8	C		13
20	Asp(O <sup>t</sup> Bu) 3- <sup>13</sup> C	--- 68261-19-8	C		13
21	Asp(OBzl) U- <sup>13</sup> C	72015-62-4 55443-54-4	D	85% enriched	15
22	Z-Cys(S-Bzl) 3- <sup>13</sup> C	79761-02-7 138307-42-3	A		17
23	Cys(S-Bzl) 1- <sup>13</sup> C	136743-61-8 ---	A		18
24	Cys(S-Bzl) 3- <sup>13</sup> C	79761-09-4 138307-42-3	A		17
25	Boc-Gln 5- <sup>15</sup> N	150464-77-0 59681-32-2	E		14
26	Boc-Gln-OBzl 5- <sup>15</sup> N	150464-75-8 59681-32-2	E		14
27	Boc-Gly <sup>15</sup> N	106665-75-2 7299-33-4	C	90% enriched	19
28	Boc-Gly <sup>15</sup> N, 1- <sup>13</sup> C <sub>2</sub>	145143-00-6 ---	G	Simultaneous protection	20,21
29	Boc-Gly <sup>15</sup> N, 1,2- <sup>13</sup> C <sub>2</sub>	145142-98-9 ---	G	Simultaneous protection	20,21
30	Boc-Gly <sup>15</sup> N, 2- <sup>13</sup> C	145143-01-7 ---	G	Simultaneous protection	20,21
31	Boc-Gly 1- <sup>13</sup> C	97352-64-2 20110-59-2	C	90% enriched	22
32	Boc-Gly U- <sup>13</sup> C	65096-45-9 67836-01-5	D	85% enriched	23
33	Boc-Gly 1,2- <sup>13</sup> C <sub>2</sub>	145142-99-0 67836-01-5	G	Simultaneous protection	20,21
34	Boc-Gly 2- <sup>13</sup> C	145143-02-8 20220-62-6	G	Simultaneous protection	20,21
35	Boc-Gly- <sup>15</sup> NH <sub>2</sub> <sup>15</sup> N, 1,2- <sup>13</sup> C <sub>2</sub>	145143-03-9 ---	G		20
36	Boc-Gly- <sup>15</sup> NH <sub>2</sub> 1,2- <sup>13</sup> C <sub>2</sub>	145143-04-0 67836-01-5	G		20
37	Boc-Gly- <sup>15</sup> NH <sub>2</sub> 2- <sup>13</sup> C	145143-05-1 20220-62-6	G		20
38	Boc-Gly-ONp <sup>15</sup> N	7299-33-4	G		24

(continued)

Table 1 continued

No.	Derivative/pattern of labelling	CAS registry no./do. free amino acid	Preparation method	Comments	Ref.(s)
39	Z-Gly 1- <sup>13</sup> C	67739-37-1 20110-59-2	C	2.25% enriched	25
40	Z-Gly 1,2- <sup>13</sup> C <sub>2</sub>	80058-03-3 67836-01-5	C		26
41	Fmoc-Gly <sup>15</sup> N	125700-33-6 7299-33-4	C		27
42	Ac-Gly <sup>15</sup> N	1449-79-2 7299-33-4	B		28
43	Ac-Gly 1- <sup>13</sup> C	--- 20110-59-2	C		29
44	Ac-Gly 2- <sup>13</sup> C	64225-28-1 20220-62-6	C		30
45	Pht-Gly 1- <sup>13</sup> C	--- 20110-59-2	G		31
46	Trt-Gly-OMe <sup>15</sup> N	98292-02-5 7299-33-4	C		32
47	Gly-OMe <sup>15</sup> N	104576-76-3 7299-33-4	C		33
48	Gly-OMe·HCl <sup>15</sup> N	72634-71-0 7299-33-4	C	60% enriched	8
49	Gly-OMe 1- <sup>13</sup> C	137248-37-4 20110-59-2	B		12
50	Gly-OMe·HCl 1,2- <sup>13</sup> C <sub>2</sub>	126614-97-9 67836-01-5	C		34
51	Gly-OEt <sup>15</sup> N	117500-42-2 7299-33-4	C		35
52	Gly-OEt·HCl <sup>15</sup> N	58420-99-8 7299-33-4	?		36
53	Gly-OEt 1- <sup>13</sup> C	143203-16-1 20110-59-2	G		37
54	Gly-OEt·HCl 1- <sup>13</sup> C	67739-40-6 20110-59-2	C	2.25% enriched	25
55	Gly-OEt·HCl 2- <sup>13</sup> C	58420-91-0 20220-62-6	?		36
56	Gly-OBzl·Tos 2- <sup>13</sup> C	114342-15-3 20220-62-6	?		38
57	BMTM-Gly-OEt <sup>a</sup> <sup>15</sup> N, 1, 2- <sup>13</sup> C <sub>2</sub>	--- ---	G		7
58	Boc-Leu <sup>15</sup> N	146953-81-3 59935-31-8	E	Simultaneous protection	39
59	Boc-Leu 1- <sup>13</sup> C	102636-64-6 74292-94-7	C		40
60	Boc-Leu <sup>15</sup> N, 1, 2- <sup>13</sup> C <sub>2</sub>	--- ---	F		7
61	Z-Leu 1- <sup>13</sup> C	125069-90-1 74292-94-7	C	90% enriched	41
62	Leu-OMe <sup>15</sup> N	--- 59935-31-8	C	90% enriched	19
63	Leu-OEt 1- <sup>13</sup> C	93523-68-3 74292-94-7	A	90% enriched; (R)?	42
64	Leu-OBzl 1- <sup>13</sup> C	115721-58-9 74292-94-7	C		40

(continued)

Table I continued

No.	Derivative/pattern of labelling	CAS registry no./do. free amino acid	Preparation method	Comments	Ref.(s)
65	Boc-Phe <sup>15</sup> N	87713-13-1 29700-34-3	C		43
66	Boc-Phe <sup>15</sup> N,1,2- <sup>13</sup> C <sub>2</sub>	---	F		7
67	Boc-Phe <sup>15</sup> N,1,2- <sup>13</sup> C <sub>2</sub> ,3,3-D <sub>2</sub>	---	F		7
68	Boc-Phe-ONp <sup>15</sup> N	29700-34-3	G		24
69	Fmoc-Phe <sup>15</sup> N	125700-32-5 29700-34-3	C		27
70	Phe-OMe <sup>15</sup> N	---	C	90% enriched	19
71	Phe-OEt <sup>15</sup> N	94601-37-3 29700-34-3	C		29
72	Z-Pro U- <sup>13</sup> C	58274-21-8 ---	D	85% enriched	44
73	Z-Pro-NH <sub>2</sub> U- <sup>13</sup> C	58274-22-9 ---	D	85% enriched	44
74	Pro-NH <sub>2</sub> U- <sup>13</sup> C	58274-20-7 ---	D	85% enriched	44
75	Pro-OMe carboxy- <sup>13</sup> C	97352-63-1 81202-06-4	C		22
76	Boc-Ser 3- <sup>13</sup> C	139623-02-2 89232-77-9	?		45
77	Fmoc-Ser 1- <sup>13</sup> C	141074-87-5 81201-84-5	C		13
78	Boc-Tyr <sup>15</sup> N	87713-11-9 35424-81-8	C		43
79	Boc-Tyr 2- <sup>13</sup> C	68882-36-0 125718-74-8	A	90% enriched	46
80	Boc-Tyr(Bzl) <sup>15</sup> N,1,2- <sup>13</sup> C <sub>2</sub>	---	F		7
81	Fmoc-Tyr <sup>15</sup> N	125700-34-7 35424-81-8	C		27
82	Tyr-OBzl-Tos 3,5- <sup>13</sup> C <sub>2</sub>	100790-33-8 70479-98-0	D		47
83	Tyr-OBzl-Tos 4- <sup>13</sup> C	100790-35-0 81201-90-3	D		47
84	Boc-Val <sup>15</sup> N	141509-91-3 59935-29-4	C		48
85	Z-Val <sup>15</sup> N	52617-00-2 59935-29-4	C		49
86	Fmoc-Val <sup>15</sup> N	125700-35-8 59935-29-4	C		27

<sup>a</sup> BMTM stands for bis(methylthio)methylene.

tional amino acid it will probably be most economical to start with a derivative with a protecting group in the sidechain. The introduction of a <sup>13</sup>C nucleus in positions 1 and/or 2 of an amino acid is more demanding. Most derivatives of this type listed in the table have been made by standard methods from

commercial amino acids. In this context it should be pointed out that relatively few authors have commented on the sterical purity of their starting materials and products. A considerable number of <sup>13</sup>C-labelled amino acids have also been made via their racemates and resolved using enzymatic meth-

ods. Even in such cases, however, the sterical purity should be checked by a sensitive, direct, preferentially chromatographic method. Otherwise, the ultimate sterical integrity of the labelled peptide product is endangered. Last but not least the challenging alternative of producing amino acids by asymmetric synthesis [50] should be mentioned. This approach has so far found limited application, but looks extremely promising [7]. It might turn out to be the method of choice for backbone-labelled amino acids, since all of the glycine isotopomers required as starting materials have already been quite easily made [20, 21].

In conclusion, it appears that the road to specifically labelled amino acids and from them to the corresponding peptides is now wide open. An extra bonus of the selective labelling approach resides in the fact that, in principle, it can be combined with the introduction of additional nuclei.

## REFERENCES

1. R. Schoenheimer and S. Ratner (1939). Studies in protein metabolism. III. Synthesis of amino acids containing isotopic nitrogen. *J. Biol. Chem.* **127**, 301–313.
2. N. S. Olsen, A. Hemingway and A. O. Nier (1943). The metabolism of glycine. I. Studies with the stable isotope of carbon. *J. Biol. Chem.* **148**, 611–618.
3. D. Shemin (1946). The biological conversion of L-serine to glycine. *J. Biol. Chem.* **162**, 297–307.
4. J. C. Reid and B. M. Tolbert in: *Amino Acids and Proteins: Theory, Methods and Applications*, D. M. Greenberg, Ed., Ch. IIIB, C. C. Thomas, Springfield, Ill. 1951.
5. R. H. Griffey and A. G. Redfield (1987). Proton-detected heteronuclear edited and correlated nuclear magnetic resonance and nuclear Overhauser effect in solution. *Quart. Rev. Biophys.* **19**, 51–82.
6. F. Degerbeck, B. Fransson, L. Grehn and U. Ragnarsson (1992). Direct synthesis of *N*-protected chiral amino acids from imidodicarbonates employing either Mitsunobu or triflate alkylation. Feasibility study using lactate with particular reference to  $^{15}\text{N}$ -labelling. *J. Chem. Soc. Perkin Trans. 1*, 245–253.
7. L. Lankiewicz, B. Nyassé, B. Fransson, L. Grehn and U. Ragnarsson (1994). Synthesis of amino acid derivatives substituted in the backbone with stable isotopes for application in peptide synthesis. *J. Chem. Soc. Perkin Trans. 1*, 2503–2510.
8. C. Niu, R. D. Bertrand, H. Shindo and J. S. Cohen (1979). Cross-peptide bond  $^{13}\text{C}$ - $^{15}\text{N}$  coupling constants by  $^{13}\text{C}$  and J cross-polarization  $^{15}\text{N}$  NMR. *J. Biochem. Biophys. Methods* **1**, 135–143.
9. B. P. Coppola, M. C. Noe, D. J. Schwartz, R. L. II Abdon and B. M. Trost (1994). Intermolecular 1,3-dipolar cycloadditions of Münchnones with acetylenic dipolarophiles: sorting out the regioselectivity. *Tetrahedron* **50**, 93–116.
10. G. B. Fields, C. G. Fields, J. Petefish, H. E. Van Wart and T. A. Cross (1988). Solid-phase peptide synthesis and solid-state NMR spectroscopy of  $[\text{Ala}^3\text{-}^{15}\text{N}][\text{Val}^1]$ -gramicidin A. *Proc. Natl Acad. Sci. USA* **85**, 1384–1388.
11. W. E. Hull and H. R. Kricheldorf (1980).  $^{15}\text{N}$ -NMR spectroscopy. 20. *Cis/trans* isomerism and neighboring residue effects of proline-containing peptides. *Biopolymers* **19**, 1103–1122.
12. B. J. Gour-Salin, P. Lachance and A. C. Storer (1991). Inhibition of papain by peptide nitriles: conversion of the nitrile group into other functionalities via the papain: nitrile thioimidate ester adduct. *Can. J. Chem.* **69**, 1288–1297.
13. T. Humbert, A. Marsura and C. Luu-Duc (1992). Solid phase synthesis of a tetrapeptide labelled with carbon-13. Preparation of L-[U- $^{13}\text{C}$ ]lysyl-L-[1- $^{13}\text{C}$ ]arginyl-L-[3- $^{13}\text{C}$ ]aspartyl-L-[1- $^{13}\text{C}$ ]serine. *J. Labelled Compd. Radiopharm.* **31**, 183–198.
14. L. Lankiewicz, L. Grehn and U. Ragnarsson (1993). Convenient preparation of Boc-L-[4- $^{15}\text{N}$ ]asparagine and -[5- $^{15}\text{N}$ ]glutamine for application in peptide synthesis. *J. Labelled Compd. Radiopharm.* **33**, 551–556.
15. F. Piriou, F. Toma, J. Savrda and S. Femandjian (1979). Synthesis and NMR analysis of cyclo-[85%  $^{13}\text{C}$ -Asp]-Pro:  $^{13}\text{C}$ - $^{13}\text{C}$  vicinal coupling constants and conformation. *Tetrahedron* **35**, 441–446.
16. H. Lam-Thanh, R. Mermet-Bouvier, S. Femandjian and P. Fromageot (1983). One-pot synthesis of 85% enriched  $^{13}\text{C}$ (U)-L-asparagine from  $^{13}\text{C}$ (U)-L-aspartic acid. *J. Labelled Compd. Radiopharm.* **20**, 143–148.
17. J. E. Baldwin, S. R. Herchen, B. L. Johnson, M. Jung, J. J. Usher and T. Wan (1981). Synthesis of  $\delta$ -(L- $\alpha$ -aminoadipoyl)-L-cysteinyl-D-valine and some carbon-13 and nitrogen-15 labelled isotopomers. *J. Chem. Soc. Perkin Trans. 1*, 2253–2257.
18. K. Uchida and M. Kainosho (1991). Synthesis of [1, 1'- $^{13}\text{C}_2$ ]-L-cystine. *J. Labelled Compd. Radiopharm.* **29**, 867–874.
19. C. Garbay-Jaureguiberry, D. Marion, E. Fellion and B. P. Roques (1982). Refinement of conformational preferences of Leu enkephalin and Tyr-Gly-Gly-Phe by  $^{15}\text{N}$  NMR. Chemical shifts and vicinal coupling constants. *Int. J. Peptide Protein Res.* **20**, 443–450.
20. L. Grehn, U. Bondesson, T. Pehk and U. Ragnarsson (1992). Synthesis of  $^{13}\text{C}$ - and  $^{15}\text{N}$ -labelled *tert*-butoxycarbonyl (Boc) glycines and glycine amides as precursors and simple models of backbone-labelled peptides. *J. Chem. Soc. Chem. Commun.*, 1332–1333.
21. L. Grehn, T. Pehk and U. Ragnarsson (1993). Synthesis and spectroscopic properties of  $^{13}\text{C}$ - and  $^{15}\text{N}$ -labelled *tert*-butoxycarbonylglycines. *Acta Chem. Scand.* **47**, 1107–1111.

22. D. W. Urry, T. L. Trapane, M. Iqbal, C. M. Venkatachalam and K. U. Prasad (1985). Carbon-13 NMR relaxation studies demonstrate an inverse temperature transition in the elastin polypeptide. *Biochemistry* 24, 5182–5189.
23. H. Lam Thanh, F. Toma, S. Fermandjian, P. Fromageot and A. Loffet in: *Peptides 1976, Proceedings of the 14th European Peptide Symposium*, A. Loffet, Ed., p. 609–616, Editions de l'Université de Bruxelles, Bruxelles 1976.
24. F. Degerbeck, B. Fransson, L. Grehn and U. Ragnarsson (1995). Synthesis of <sup>15</sup>N-labelled Leu-enkephalins. *Acta Chem. Scand.* 49, 149–151.
25. M. Hollosi, Z. Dobolyi and G. Bujtas (1978). Syntheses of cyclic peptides from glycine partially labelled with deuterium and/or carbon-13 and two pairs of proline. *Liebigs Ann. Chem.*, 796–803.
26. J. Schaefer, T. A. Skokut, E. O. Stejskal, R. A. McKay and J. E. Varner (1981). Estimation of protein turnover in soybean leaves using magic angle double cross-polarization nitrogen-15 nuclear magnetic resonance. *J. Biol. Chem.* 256, 11574–11579.
27. C. G. Fields, G. B. Fields, R. L. Noble and T. A. Cross (1989). Solid phase peptide synthesis of <sup>15</sup>N-gramicidins A, B, and C and high performance liquid chromatographic purification. *Int. J. Pept. Protein Res.* 33, 298–303.
28. J. W. Reed and D. G. I. Kingston (1986). Biosynthesis of antibiotics of the virginiamycin family. 5. The conversion of phenylalanine to phenylglycine in the biosynthesis of virginiamycin S<sub>1</sub>. *J. Nat. Prod.* 49, 626–630.
29. T. G. Oas, C. J. Hartzell, T. J. McMahon, G. P. Drobny and F. W. Dahlquist (1987). The carbonyl <sup>13</sup>C chemical shift tensors of five peptides determined from <sup>15</sup>N dipole-coupled chemical shift powder patterns. *J. Am. Chem. Soc.* 109, 5956–5962.
30. J. L. C. Wright, L. C. Vining, A. G. McInnes, D. G. Smith and J. A. Walter (1977). Use of <sup>13</sup>C in biosynthetic studies. The labelling pattern in tenellin enriched from isotope-labelled acetate, methionine, and phenylalanine. *Can. J. Biochem.* 55, 678–685.
31. K. Kurumaya, T. Okazaki, N. Seido, Y. Akasaka, Y. Kawajiri, M. Kajiwara and M. Kondo (1989). A facile synthesis of δ-aminolevulinic acid (ALA) regioselectively labeled with <sup>13</sup>C and direct observation of enzymatic transformation from ALA to porphobilinogen. *J. Labelled Compd. Radiopharm.* 27, 217–235.
32. S. Akoka, C. Meir, C. Tellier, C. Belaud and S. Poignant (1985). Synthesis of nitrogen-15 enriched phospholipids by transphosphatidylation. *Synth. Commun.* 15, 101–107.
33. S. Akoka, C. Tellier and S. Poignant (1986). Molecular order, dynamics, and ionization state of phosphatidylethanolamine bilayers as studied by <sup>15</sup>N NMR. *Biochemistry* 25, 6972–6977.
34. P. C. Myhre, C. T. Maxey, D. C. Bebout, S. H. Swedberg and B. L. Petersen (1990). Precursors to carbon-13 labelled reactive intermediates: preparation and NMR characterization of two double label isomers of methyl cyclopropene-3-carboxylate. *J. Org. Chem.* 55, 3417–3421.
35. S. M. Ludeman, E. M. Shulman-Roskes, M. P. Gamcsik, T. G. Hamill, Y. H. Chang, K. I. Koo and O. M. Colvin (1993). Synthesis of <sup>15</sup>N and <sup>17</sup>O labelled phosphoramidate mustards. *J. Labelled Compd. Radiopharm.* 33, 313–326.
36. R. L. Ellsworth, H. E. Mertel and W. J. A. VandenHeuvel (1976). Synthesis and mass spectrometry of isotopically labelled isopropyl 2-(4-thiazolyl)-5-benzimidazolecarbamate (cambendazole). *J. Agric. Food Chem.* 24, 544–549.
37. H. Duplaa, O. Convert, A.-M. Sautereau, J.-F. Tocanne and G. Chassaing (1992). Binding of substance P to monolayers and vesicles made of phosphatidylcholine and/or phosphatidylserine. *Biochim. Biophys. Acta* 1107, 12–22.
38. S. W. Elson, S. R. Woroniecki and K. H. Baggaley. Preparation of clavulanic acid precursors. Eur. Pat. Appl. EP 213,914 [C.A. 108 (1988) 204405u].
39. F. Degerbeck, B. Fransson, L. Grehn and U. Ragnarsson (1993). Synthesis of <sup>15</sup>N-labelled chiral Boc-amino acids from triflates: enantiomers of leucine and phenylalanine. *J. Chem. Soc. Perkin Trans. 1*, 11–14.
40. M. K. Lutek, T. Hofmann and C. M. Deber (1988). Transpeptidation reactions of porcine pepsin. Formation of tetrapeptides from dipeptide substrates. *J. Biol. Chem.* 263, 8011–8016.
41. B. Danieli, F. M. Rubino and A. Cremonesi (1989). Fast atom bombardment mass spectrometry of carbobenzyloxy-protected amino acids and peptides. *Org. Mass. Spectrom.* 24, 225–229.
42. S.-S. Yuan and A. M. Ajami (1984). Stable isotope labelled alpha-keto acids. *J. Labelled Compd. Radiopharm.* 21, 735–742.
43. D. Cowburn, D. H. Live, A. J. Fischman and W. C. Agosta (1983). Side chain conformations of oxytocin and vasopressin studied by NMR observation of isotopic isomers. *J. Am. Chem. Soc.* 105, 7435–7442.
44. W. Haar, S. Fermandjian, J. Vicar, K. Blaha and P. Fromageot (1975). <sup>13</sup>C-nuclear magnetic resonance study of [85% <sup>13</sup>C-enriched proline]thyrotropin releasing factor: <sup>13</sup>C-<sup>13</sup>C vicinal coupling constants and conformation of the proline residue. *Proc. Natl Acad. Sci. USA* 72, 4948–4952.
45. S. N. Ludwig and C. J. Unkefer (1992). Stereoselective synthesis of stable isotope labelled L-α-amino acids: synthesis of L-[4-<sup>13</sup>C] and L-[3,4-<sup>13</sup>C]aspartic acid. *J. Labelled Compd. Radiopharm.* 31, 95–102.
46. M. Blumenstein, V. J. Hruby and D. M. Yamamoto (1978). Evidence from hydrogen-1 and carbon-13 nuclear magnetic resonance studies that the dissociation rate of oxytocin from bovine neurophysin at neutral pH is slow. *Biochemistry* 17, 4971–4977.
47. T. E. Walker, C. Matheny, C. B. Storm and H. Hayden

- (1986). An efficient chemomicrobiological synthesis of stable isotope-labelled L-tyrosine and L-phenylalanine. *J. Org. Chem.* 51, 1175–1179.
48. S. M. Holl, G. R. Marshall, D. D. Beusen, K. Kociolek, A. S. Redlinski, M. T. Leplawy, R. A. McKay, S. Vega and J. Schaefer (1992). Determination of an 8-Å interatomic distance in a helical peptide by solid-state NMR spectroscopy. *J. Am. Chem. Soc.* 114, 4830–4833.
49. S. John and D. Gröger (1974). Synthesen von radioaktiven potentiellen Vorstufen des Peptidteiles von Ergotoxin-Alkaloiden. *Pharmazie* 29, 181–186.
50. R. M. Williams: *Synthesis of Optically Active  $\alpha$ -Amino Acids*, Pergamon Press, Oxford 1989.